Occurrence of adenovirus infection and clinical impact in paediatric stem cell transplant recipients

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Summary

In this study, the occurrence and clinical impact of adenovirus (AdV) infection was investigated in paediatric hematopoietic stem cell transplantation (HSCT) recipients. A number of 603 specimens (including whole blood, respiratory and other samples) from 181 patients were tested by real-time polymerase chain reaction; clinical outcome was investigated. Overall, 118/603 (19.6%) specimens from 21/181 (11.6%) patients resulted positive to AdV (including 17.3, 29.9, 17.6, and 15.8% of total number of whole blood, respiratory, urine and other specimens, respectively). On whole blood specimens, viral loads ranged from <600 (limit of detection) to >5×10^6 copies/mL, with a median value 2×10^4. Multiple specimens were positive in patients in which viral load on whole blood was high. Adenoviral positivity on whole blood was associated to poor prognosis, as death occurred in three of ten (30%) patients with persistent positivity on whole blood specimens, also despite the administration of an antiviral agent (cidofovir).

Introduction

Hematopoietic stem cell transplantation (HSCT) represents the last treatment for a variety of diseases, including lymph proliferative disorders, leukaemia, and bone marrow failure syndromes. However, transplant-related mortality remains the main limitation to its success. In addition to the development of graft-versus-host disease (GVHD), infections are important complications during HSCT procedures and contribute significantly to morbidity and transplantation-related mortality. The periods at highest risk of infections occur during the immunosuppressed phase and in the period preceding the immune reconstitution. The risk of infection is higher for allogeneic HSCT, GVHD, delayed immune reconstitution and the incidence and severity of infections strongly correlate with pretransplantation serostatus, source of HSC, time elapsed for HSCT, unrelated of human leukocyte antigen-mismatched related donor transplantation, immunosuppressive therapy, use of total body irradiation, fludarabine, antilymphocyte serum and monoclonal antibodies during the conditioning regimen (8,13).

Viral infections/reactivations represent one of the main infectious risk in HSCT (7,10,12). The viral agents mainly involved are cytomegalovirus, Epstein-Barr virus, adenovirus (AdV), varicella zoster virus and herpes simplex virus. In the last years, there has been an increase in the number of viral diseases observed in these patients. This may be at least partially due to an improvement in diagnostic facilities, but the increasing number of transplants procedures and the more severe immunosuppression may also have played an important role. Recently, AdV has been increasingly recognised with emerging significance after transplantation, with a particularly aggressive course of viral disease (e.g. severe respiratory disease, hepatitis, colitis, haemorrhagic cystitis, and keratoconjunctivitis), and a higher mortality rate (8-26%). The frequency of AdV infections in paediatric transplant recipients is significantly higher than in adults. About 2-5 fold higher infection rates have been reported (1-6,10-14). Young paediatric transplant recipients seem to be at especially high risk, possibly because of a higher rate of primary infection as compared to reactivation in adolescent patients or adults. Patient selection for antiviral therapy, choice of drug, optimal timing of therapy and monitoring of therapeutic effects, treatment of AdV infections remain a domain of controversy. A recent article by Minarek and colleagues (9) found that despite a low mortality directly attributed to AdV infection, blood AdV loads >10,000 copies/mL were significant and independent risk factors.
for poor survival and recommended AdV blood monitoring for at least 50 days for paediatric HSCT with early occurrence of DNA positivity in blood predisposing for development of high viral loads.

Materials and Methods

In this study, the occurrence and clinical impact of AdV infection was investigated in a population of paediatric HSCT recipients. Of patients 181 (106 males/75 females; mean age, 8.4±5.6 years; range, 5-19) who underwent HSCT were evaluated for AdV infection at the Microbiology and Virology Unit of the University Hospital Città della Salute e della Scienza di Torino (Turin, Italy).

AdV was not tested routinely, but in the presence of clinical and/or laboratory and/or radiological signs and/or symptoms suggestive of AdV infection (i.e. systemic symptoms, including fever, and organ-specific symptoms indicative of respiratory, keratoconjunctival, gastrointestinal, urinary, central nervous system involvement) or for checking up a previous positivity (in this case usually at weekly until negativity).

Overall, 603 specimens collected on clinical indication (including occurrence of systemic or organ disease) were evaluated: in particular, 472 whole blood specimens from 114 patients; 87 respiratory specimens (nasopharyngeal swabs, bronchoaspirate, bronchoalveolar lavage) from 66 patients; 17 urine samples from nine patients; 8 keratoconjunctival swabs from two patients; and 19 other specimens (including cerebrospinal fluid, bone marrow, pleural and peritonitic fluid, tissue biopsy). Multiple and different specimens were available for 56 patients (mainly whole blood and one or more other samples).

All the specimens were tested by a commercially available real-time polymerase chain reaction (PCR) assay (Adenovirus ELItel MGB® kit; ELITech Italy, Trezzano s.N., Italy) following automated extraction by the Qiashympson instrument (Qiagen, Hilden, Germany). Amplification was performed with the 7500 real-time PCR systems (Applied Biosystems, Foster City, CA, USA).

Results

Overall, 118/603 (19.6%) specimens from 21/181 (11.6%) patients resulted positive to AdV. In particular, considering the different specimens, 82/472 (17.3%) whole blood specimens from 10/114 (8.7%) patients for which this specimen was tested; 26/87 (29.9%) respiratory samples from 11/66 (16.7%) patients; three of 17 (17.6%) urine specimens from 2/9 (22.2%) patients; and three of 19 (15.8%) other specimens from two patients. No keratoconjunctival sample was positive. On whole blood specimens, viral loads ranged from <600 (limit of detection) to >5×10⁶ copies/mL, with a median value 2×10⁴; in particular, this was <600 copies/mL in 14/82 (17.1%) specimens, between 600 and 10,000 copies/mL in 12 (14.6%), and >10,000 copies/mL in the remaining samples. Peak viral loads were <600 in 2/10 (20%) patients, between 600 and 10,000 in two (20%), and >10,000 in six (60%). Multiple specimens were positive in patients in which viral load on whole blood was high. Adenoviral positivity on whole blood was associated to poor prognosis, as death occurred in three of ten (30%) patients with persistent positivity on whole blood specimens, in the absence of other infections or progression of underlying disease. In two of these patients (a 16 year-old girl and a 9-year-old boy) viral load values >10⁴ copies/mL whole blood persisted for 3 and 6 months, respectively (until death), despite the administration of an antiviral agent (cidofovir), suggesting that both high viral load and persistent positivity could be suggestive of poor prognosis. Patients’ features in relation to viral load in whole blood specimens are reported in Table 1.

Conclusions

Adenovirus has been increasingly recognized as possibly playing a relevant role in clinical outcome of HSCT paediatric patients. Based on the results of this study, AdV infection can account for systemic and/or organ-specific signs/symptoms in approximately 10% of paediatric HSCT recipients. A wide range of occurrence rates of organ or systemic AdV infection in HSCT patients has been reported in previous studies (1-7,10-14). Risk factors associated to AdV infection in HSCT include T-cell depletion, unrelated and cord blood HSCT, GVHD, and lymphopenia. Based on our and other studies, AdV infections appears to play a relevant role in terms of patient outcome, particularly in the presence of disseminated infection, that usually follows a primary local infection. AdV positivity can be detected in different clinical specimens, with high and persisting viral load on whole blood specimens. In a previous study (9), early detection of AdV in blood before day 50 post transplantation would improve survival. AdV monitoring may be useful for the detection and early treatment of this virus.
was a major independent risk factor for the development of high viral loads (>10,000 copies/mL). In these patients, pre-emptive cidofovir treatment correlated with low AdV-attributed mortality. At moment, there is no indication for routine monitoring of AdV in paediatric HSCT recipients; however, AdV aetiology of infectious transplant complications should be taken in account. Further studies are necessary to identify type, dosage and timing of antiviral treatment, and the potential role for AdV-specific T-cell therapy (8).

References